GRADUATE PERMISSION TO CROSS-LIST FORM

This form must ALWAYS be accompanied by a graduate COURSE FORM.

Contact Name: Christine Byrum     Email: byrumc@cofc.edu     Phone:  (893)953-5504

Department and School Name: Biology, School of Sciences and Mathematics
Name and Acronym of Graduate Program: Marine Biology Program, MS

Date (Semester/Year) cross-listing will take effect: Fall 2015

I. CATEGORY OF REVIEW (Check all that apply)
   ☑ New Course -- Course Number/Title BIOL 623/623L (Genomics/Genomics Lab)
   ☐ Existing Course -- Course Number/Title
   ☐ Special Topic Course -- Course Number/Title

   This course will be cross-listed with an
   ☑ undergraduate course (complete sections II, III, and IV below)
   ☐ existing graduate course (complete section IV below)

II. CURRICULUM DIFFERENCES –UNDERGRADUATE AND GRADUATE LEVELS
    Please submit separate syllabi for both graduate and undergraduate courses

   Syllabi for both undergraduate and graduate courses are attached ☑ YES ☐ NO

   Explain the differences between the syllabi in terms of requirements, learning objectives and course content

   The graduate version of the course (BIOL 623/623L) differs from the undergraduate version (BIOL 423/423L) in that the graduate students are expected to lead an additional discussion session and to write an additional paper for the class.
III. APPROVAL SECTION – GRADUATE COURSE WITH UNDERGRADUATE COURSE

Undergraduate Course Number / Title 423/423L (Genomics/Genomics Lab)

Names and Signatures:

Name of Department Chair of the Graduate Course Jaap Hillenius
Signature ___________________________ Date: 1/28/15

Department Chair of the Undergraduate Course Jaap Hillenius
Signature ___________________________ Date: 1/28/15

Graduate Program Director Craig Plante
Signature ___________________________ Date: 1/28/15

Provost Brian McGee
Signature ___________________________ Date: 

IV. APPROVAL SECTION – GRADUATE COURSE WITH EXISTING GRADUATE COURSE

Graduate Course Number / Title of Existing Graduate Course

Program(s) of Existing Graduate Course

Names and Signatures:

“Host” Program Director
Signature ___________________________ Date: 

“Requesting” Program Director
Signature ___________________________ Date: 

Provost
Signature ___________________________ Date: 

September 2011
Return form to the Graduate School Office for Further Processing

Signature of Chair of the Faculty Committee on Graduate Education, Continuing Education & Special Programs:

[Signature]

Date: 4/28/15

Signature of Chair of the Graduate Council:

[Signature]

Date: 

Signature of Faculty Senate Secretary:

[Signature]

Date: 

Date Approved by Faculty Senate: 

[Signature]
Biology 423/423L
Genomics Lecture/Lab

Lectures: MF 11:00 am-12:15 pm, New Science Center, Room 140
Instructor: Dr. Christine Byrum
Email: byrumc@cofc.edu
Phone: (843) 953-7176
Office: 236B RHSC, College of Charleston (Downtown campus)
Office Hours: By appointment

Course Overview:
This upper level undergraduate course is designed to familiarize students with the field of genomics and current topics of interest in this quickly expanding discipline. We will explore several subdisciplines in the field and learn about a variety of techniques applicable to genomic analysis. Particular attention will be devoted to new advances in the area of marine genomics as well as biomedically relevant areas of genomics. Materials will be presented as lectures, classroom discussions of journal articles, and labs/workshops.

Objectives:
Course goals include the following:
1) Students should become familiar with terminology used by genomicists as well as key principles and theories associated with this field.
2) They will be expected to coherently explain genomic concepts and to apply these concepts to new situations or to predict outcomes based on what they have learned in class.
3) Students will learn how to use bioinformatic tools to find/identify genes in a genome and to better characterize gene products.
4) Become familiar with recent genomic research through discussion of the scientific literature.
5) Learn techniques in phylogenetic analysis that are applicable to genomic studies.
6) Become familiar with all aspects of qPCR analysis including sample preparation, primer design, primer testing, calibration of the qPCR machine, controls necessary for qPCR analysis, running qPCR reactions, and analysis of qPCR results.
7) Learn about sequencing technologies, gene expression analysis, and proteomics.
8) Discuss ethical issues relevant to genomics.
9) Become more aware of local genomic resources.
10) Learn how to present genomic information in a talk and/or a paper.
Optional Texts:

Prerequisites: Genetics (BIOL 305), Introduction to Statistics 1 (MATH 250), Molecular Biology (BIOL 312) / Cell Biology (BIOL 313) or permission of the instructor.

**Course Policies**

**Attendance:** Regular classroom participation is critical in this course. If the student is unable to attend a class, he/she should be sure to get the information from a classmate or from the instructor so that he/she doesn’t fall behind. If an absence is anticipated, the instructor should be informed ahead of time.

**Class Assignments:**

**Quizzes:** There will be seven quizzes during the course of the semester at which the students will be assessed to determine their knowledge concerning recent and/or previous materials. These will typically consist of five to ten short answer/essay questions. This is a significant portion of your grade and you should be sure to prepare thoroughly for each evaluation.

**Discussions:** Students will also lead class discussions about topics in genomics. This will be done in teams of two. Students should introduce the class to the topic and lead a discussion reviewing two papers (30 minutes/person). Students should prepare typed notes for these discussions (notes highlighting key points in the introduction and listing discussion questions with written answers.). These notes will be handed in following the discussion session.

Those not presenting are expected to read the assigned papers before class and to participate in class discussions. Participation grades are based on the following: A) Did the student take the time to carefully read this paper? Is he/she able to answer questions raised during discussions? Can he/she describe what was done in the paper? Has he/she thought about implications of this work? B) Is the student willing to participate and contribute to classroom discussions? Even if you are shy, it is important that you make an effort to actively participate. C) Has the student taken any extra steps to build on what they’ve learned after reading the paper? Does he/she contribute extra information in these discussions?

**Out of Class Exercises:** These are short “homework” assignments. Often these are worksheets to fill out or exercises to do that will improve understanding of a technique previously discussed in class or lab. Students are encouraged to talk to each other about these assignments but should not copy work. Make sure that all work is shown and turn in assignments at the
beginning of the next class period. Assignments will not be accepted after all papers have been corrected and turning in an assignment late may result in a penalty.

**Class Projects:** Over the semester, students will work together to complete a course project in the lab during which they analyze expression of a set of genes in the sea urchin. Students will find members of a gene family, identify domains present in the members of that gene family, identify human genes that are likely homologues based on gene alignments and perform phylogenetic analysis to characterize the evolutionary relationships of these genes to those in other phyla. They will also design primers and perform QPCR analysis to determine levels of gene expression at different developmental stages. Each individual will submit his/her project as a paper.

**Grading of Assignments:** A single grade based on performance in both the lecture and lab will be assigned for the 4-credit Genomics course. The following criteria will be used to calculate the grade.

- **Quizzes:** 30%
- **Participation in Class Discussions (lecture):** 10%
- **Leading Class Discussions in Team of Two (2) (lecture):** 20%
- **Out of Class Exercises (lab/lecture):** 20%
- **Class Project (lab):** 20%

**Grading Scale:**
- 93 and above: A
- 90-92.9: A-
- 87-89.9: B+
- 83-86.9: B
- 80-82.9: B-
- 77-79.9: C+
- 73-76.9: C
- 70-72.9: C-
- 67-69.9: D+
- 63-66.9: D
- 60-62.9: D-
- below 60: F

**Classroom courtesy:** Students are expected to turn off cell phones, beepers, and any other disruptive devices during lectures and discussions. Exceptions will be made in extreme situations such as spouses anticipating the birth of a child or a serious emergency. Permission to leave an electronic device on should be obtained prior to class.

**Academic Integrity:** Students are expected to behave in an honest and responsible manner. Violations of the honor code are offensive and will generally be dealt with severely. We will adhere to the following policy as quoted from the Honor Council's recommended guidelines:

"Lying, cheating, attempted cheating, and plagiarism are violations of our Honor Code that, when identified, are investigated. Each incident will be examined to determine the degree of deception involved."
In cases where it is determined that a student’s actions were due to a misunderstanding, the situation will be handled by the instructor. A written intervention designed to help prevent the student from repeating the error will be given to the student. The intervention, submitted by form and signed by both the instructor and the student, will be forwarded to the Dean of Students and placed in the student’s file. Cases of suspected academic dishonesty will be reported directly by the instructor and/or others having knowledge of the incident to the Dean of Students. A student found responsible by the Honor Board for academic dishonesty will receive a XF in the course, indicating failure of the course due to academic dishonesty. This grade will appear on the student’s transcript for two years after which the student may petition for the X to be expunged. The student may also be placed on disciplinary probation, suspended (temporary removal) or expelled (permanent removal) from the College by the Honor Board.

Students should be aware that unauthorized collaboration--working together without permission--is a form of cheating. Unless the instructor specifies that students can work together on an assignment, quiz and/or test, no collaboration during the completion of the assignment is permitted. Other forms of cheating include possessing or using an unauthorized study aid (which could include accessing information stored on a cell phone), copying from others’ exams, fabricating data, and giving unauthorized assistance.

Research conducted and/or papers written for other classes cannot be used in whole or in part for any assignment in this class without obtaining prior permission from the instructor. Students can find the complete Honor Code and all related processes in the Student Handbook at http://www.cofc.edu/generaldocuments/handbook.pdf.”

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**Lecture Schedule**

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<td>Aug. 26 M</td>
<td>Review of Molecular Techniques (Ch. 2)</td>
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<td>Aug. 30 F</td>
<td>Discussion - Origins of Genomics (Assigned Reading) – Concepts Test 1</td>
</tr>
<tr>
<td>Sept. 2 M</td>
<td>Sequencing Genomes (Ch. 7, Ch. 17)</td>
</tr>
<tr>
<td>Sept. 6 F</td>
<td>Discussion - Genome Sequencing Technologies</td>
</tr>
<tr>
<td>Sept. 9 M</td>
<td>Gene Finding/Annotation (Assigned Reading)</td>
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</table>
Sept. 13 F  Discussion – Alternative Splicing – Concepts Test 2
Sept. 16 M  Measuring Gene Expression (Ch. 20)
Sept. 20 F  Discussion - EST projects
Sept. 23 M  Personalized Medicine
Sept. 27 F  Discussion – Personalized Medicine – Concepts Test 3
Sept. 30 M  QPCR (Assigned Reading)
Oct. 4 F    Discussion – Gene Expression
Oct. 7 M    Proteomics (Ch. 21-23)
Oct. 11 F   Discussion - Proteomics/Metabolomics – Concepts Test 4
Oct. 14 M   Fall Break
Oct. 18 F   Microbial Genomics (Assigned Reading)
Oct. 21 M   Discussion – Microbial Genomics
Oct. 25 F   Studying Genome Variation (Ch. 25)
Oct. 28 M   Comparative Genomics (Ch. 18)
Nov. 1 F    Discussion – Comparative Genomics – Concepts Test 5
Nov. 4 M    Sequence Alignment (Ch. 9, Assigned Reading)
Nov. 8 F    Discussion – RNAi
Nov. 11 M   Phylogenetic Analysis (Assigned Reading)
Nov. 15 F   Discussion - Ethical Issues
Nov. 18 M   Discussion – Epigenetics – Concepts Test 6
Nov. 22 F   Functional Genomics (Ch. 19)
Nov. 25 M   Discussion – Functional Genomics
Genomics Lab

Objectives of the Genomics Lab: In the Genomics Lab, a mixture of computer-based labs and “wet lab” exercises are used to familiarize students with key procedures regularly used to study genomics. We will work together in teams to ask real scientific questions. Students will learn how to:

1) Retrieve nucleotide (transcriptomic and genomic DNA) and protein sequences from scientific databases.
2) Perform simple and advanced BLAST searches.
3) Design standard and QPCR primer sets for reverse transcriptase PCR and QPCR.
4) Extract RNA from an organism and produce cDNA.
5) Perform and analyze the results of reverse transcriptase PCR and QPCR.
6) Extract DNA from a gel after electrophoresis and prepare samples for sequencing.
7) Analyze sequence data.
8) Find homologous genes for phylogenetic analysis.
9) Perform phylogenetic analyses using MEGAS.

Lab Safety and Attire: Before attending the first lab, each student should review the official SSM safety manual posted on OAKS. Many of the labs are computer-based, but others involve the use of hazardous chemicals. During “wet labs”, students should dress appropriately, wearing closed-toed shoes and pants rather than shorts. These protect more in the case of a chemical mishap.
# Genomics Lab Schedule

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| Aug. 30 | BLAST lab  
 _**Goal:**_ Become proficient at performing different types of BLAST searches on NCBI's BLAST site. Learn how to select the correct BLAST program for your question, which databases are available on BLAST, parameters available, and how to perform specialized BLAST searches.  
 _**Assignment:**_ Complete BLAST worksheet before next lab. |
| Sept. 6 | Finding Genes in a Genome/Domain Analysis  
 _**Goal:**_ Outline lab organization, goals for semester. Learn to search for annotated and unannotated genes. Discuss how to deal with “problematic” sequences. Learn to do domain searches (Pfam, NCBI, SMART, TMHMM) and how to make drawings showing key domains.  
 _**Assignment:**_ Practice finding genes and characterizing domain structure in these products. |
| Sept. 13 | Standard and QPCR Primer Design/Review of PCR  
 _**Goal:**_ Learn how to make primers for standard PCR and QPCR analysis.  
 _**Assignment:**_ Design Standard PCR and QPCR primer sets to be used later this semester. |
| Sept. 20 | Collecting Specimens for QPCR Analysis  
 _**Goal:**_ Learn how to prepare specimens for QPCR analysis. Induce spawning in sea urchins, culture and collect embryos at specific developmental stages, and prepare for RNA extraction.  
 _**Assignment:**_ Obtain specimens for QPCR labs. |
| Sept. 27 | Extracting RNA/Producing cDNA  
 _**Goal:**_ Collect/extract RNA and use reverse transcriptase to produce cDNA for RT-PCR or QPCR.  
 _**Assignment:**_ Isolate mRNA and produce cDNA. |
| Oct. 4  | Performing the Standard PCR Reaction  
 _**Goal:**_ Test the standard primers that you made using a standard PCR reaction.  
 _**Assignment:**_ Obtain PCR products (if possible) using your primer sets before the next lab. |
| Oct. 11 | Electrophoresis of PCR products/Gel Extractions for Sequencing  
 _**Goal:**_ Evaluate PCR products obtained in last lab using gel electrophoresis and prepare samples for sequencing. |
Assignment: Determine whether primers amplified genes of interest and verify by sequencing the product.

Oct. 18
Tour of Genome Core Facilities at Fort Johnson
Goal: Become familiar with technologies available nearby for RNA Seq, microarray studies, and metabolomic analysis.
Assignment: Continue to work on class project paper.

Oct. 25
Optimizing for QPCR/Running Reactions
Goal: Run a QPCR reaction after performing necessary tests.
Assignment: Acquire QPCR data.

Nov. 1
Optimizing for QPCR/Running Reactions
Goal: Continue collecting data for class project.
Assignment: Acquire QPCR data.

Nov. 8
Analyzing QPCR data/ Finding Potential Homologues
Goal: Complete QPCR analysis for class project and related analysis. Also, introduce websites useful for finding homologous genes (Ensembl, Homologene, HUGO, Mouse Genome Informatics, etc.).
Assignment: Analyze QPCR data.

Nov. 15
Sequence Alignment
Goal: Learn how to do pairwise and multiple sequence alignments. Discuss editing multiple sequence alignments.
Assignment: Perform pairwise and multiple sequence alignments on your project genes to prepare for phylogenetic analysis. Edit the multiple sequence alignment.

Nov. 22
Generating Phylogenies to Identify Homologues
Goal: Learn to generate neighbor-joining and maximal parsimony trees in MEGA.
Assignment: Produce a neighbor-joining tree and a maximal parsimony tree for a family of genes you are working on. Compare your genes to potential homologues in humans. Then perform a second analysis in which the genes are compared to a range of organisms to assess evolution of the gene.

* Schedule may vary subject to scheduling changes and other modifications as needed.

Bold dates: Lab will be held in RHSC 315
Biology 623/623L
Genomics Lecture/Lab

Lectures: MF 11:00 am-12:15 pm, New Science Center, Room 140
Instructor: Dr. Christine Byrum
Email: byr umc@cofc.edu
Phone: (843) 953-7176
Office: 236B RHSC, College of Charleston (Downtown campus)
Office Hours: By appointment

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4) Become familiar with recent genomic research through discussion of the scientific literature.
5) Learn techniques in phylogenetic analysis that are applicable to genomic studies.
6) Become familiar with all aspects of QPCR analysis including sample preparation, primer design, primer testing, calibration of the QPCR machine, controls necessary for QPCR analysis, running QPCR reactions, and analysis of QPCR results.
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Optional Texts:

Prerequisites: Genetics (BIOL 305), Molecular Biology (BIOL 312) / Cell Biology (BIOL 313) or permission of the instructor.

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Solo Discussion: In addition to the two team-led discussions, each graduate student will be expected to lead a third discussion session independently. He/she will select 1-2 papers to discuss and will prepare a Powerpoint presentation (20-25 minutes) to present before the
discussion that will give the other students some background on the topic. Following this introduction, the student will lead a classroom discussion about the chosen paper(s).

**Out of Class Exercises:** These are short “homework” assignments. Often these are worksheets to fill out or exercises to do that will improve understanding of a technique previously discussed in class or lab. Students are encouraged to talk to each other about these assignments but should not copy work. Make sure that all work is shown and turn in assignments at the beginning of the next class period. Assignments will not be accepted after all papers have been corrected and turning in an assignment late may result in a penalty.

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<thead>
<tr>
<th>Category</th>
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</tr>
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<tbody>
<tr>
<td>Tests</td>
<td>30%</td>
</tr>
<tr>
<td>Participation in Class Discussions (lecture)</td>
<td>10%</td>
</tr>
<tr>
<td>Leading Class Discussions in Team of Two (2) (lecture)</td>
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<td>Solo Class Discussion (1) (lecture)</td>
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<tr>
<td>Out of Class Exercises (lab/lecture)</td>
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<td>Class Project (lab)</td>
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<td>QPCR (Assigned Reading)</td>
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<td>Oct. 4 F</td>
<td><em>Discussion</em> – Gene Expression</td>
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<td>Oct. 7 M</td>
<td>Proteomics (Ch. 21-23)</td>
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<td>Oct. 11 F</td>
<td><em>Discussion</em> - Proteomics/Metabolomics – Concepts Test 4</td>
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<tr>
<td>Oct. 14 M</td>
<td>Fall Break</td>
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<td>Oct. 18 F</td>
<td>Microbial Genomics (Assigned Reading)</td>
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Nov. 1 F  Discussion – Comparative Genomics – Concepts Test 5
Nov. 4 M  Sequence Alignment (Ch. 9, Assigned Reading)
Nov. 8 F  Discussion – RNAi
Nov. 11 M  Phylogenetic Analysis (Assigned Reading)
Nov. 15 F  Discussion - Ethical Issues
Nov. 18 M  Discussion – Epigenetics – Concepts Test 6
Nov. 22 F  Functional Genomics (Ch. 19)
Nov. 25 M  Discussion – Functional Genomics
Nov. 29 F  Thanksgiving
Dec. 2 M  Discussion/Presentation of Results from Lab Projects – Concepts Test 7

* Schedule may vary subject to scheduling changes and other modifications as needed.

**Objectives of the Genomics Lab:** In the Genomics Lab, a mixture of computer-based labs and “wet lab” exercises are used to familiarize students with key procedures regularly used to study genomics. We will work together in teams to ask real scientific questions. Students will learn how to:

1) Retrieve nucleotide (transcriptomic and genomic DNA) and protein sequences from scientific databases.
2) Perform simple and advanced BLAST searches.
3) Design standard and QPCR primer sets for reverse transcriptase PCR and QPCR.
4) Extract RNA from an organism and produce cDNA.
5) Perform and analyze the results of reverse transcriptase PCR and QPCR.
6) Extract DNA from a gel after electrophoresis and prepare samples for sequencing.
7) Analyze sequence data.
8) Find homologous genes for phylogenetic analysis.
9) Perform phylogenetic analyses using MEGA5.

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wearing pants rather than shorts and closed-toed shoes. These protect more in the case of a chemical mishap.

## Genomics Lab Schedule

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<td>Aug. 30</td>
<td>BLAST lab</td>
<td><em>Goal:</em> Become proficient at performing different types of BLAST searches on NCBI’s BLAST site. Learn how to select the correct BLAST program for your question, which databases are available on BLAST, parameters available, and how to perform specialized BLAST searches. <em>Assignment:</em> Complete BLAST worksheet before next lab.</td>
</tr>
<tr>
<td>Sept. 6</td>
<td>Finding Genes in a Genome/Domain Analysis</td>
<td><em>Goal:</em> Outline lab organization, goals for semester. Learn to search for annotated and unannotated genes. Discuss how to deal with “problematic” sequences. Learn to do domain searches (Pfam, NCBI, SMART, TMHMM) and how to make drawings showing key domains. <em>Assignment:</em> Practice finding genes and characterizing domain structure in these products.</td>
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<tr>
<td>Sept. 13</td>
<td>Standard and QPCR Primer Design/Review of PCR</td>
<td><em>Goal:</em> Learn how to make primers for standard PCR and QPCR analysis. <em>Assignment:</em> Design Standard PCR and QPCR primer sets to be used later this semester.</td>
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<tr>
<td>Sept. 20</td>
<td>Collecting Specimens for QPCR Analysis</td>
<td><em>Goal:</em> Learn how to prepare specimens for QPCR analysis. Induce spawning in sea urchins, culture and collect embryos at specific developmental stages, and prepare for RNA extraction. <em>Assignment:</em> Obtain specimens for QPCR labs.</td>
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<tr>
<td>Sept. 27</td>
<td>Extracting RNA/Producing cDNA</td>
<td><em>Goal:</em> Collect/extract RNA and use reverse transcriptase to produce cDNA for RT-PCR or QPCR. <em>Assignment:</em> Isolate mRNA and produce cDNA.</td>
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<td>Oct. 4</td>
<td>Performing the Standard PCR Reaction</td>
<td><em>Goal:</em> Test the standard primers that you made using a standard PCR reaction. <em>Assignment:</em> Obtain PCR products (if possible) using your primer sets before the next lab.</td>
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</table>
Oct. 11  Electrophoresis of PCR products/Gel Extractions for Sequencing  
  *Goal*: Evaluate PCR products obtained in last lab using gel electrophoresis and prepare samples for sequencing.  
  *Assignment*: Determine whether primers amplified genes of interest and verify by sequencing the product.

Oct. 18  Tour of Genome Core Facilities at Fort Johnson  
  *Goal*: Become familiar with technologies available nearby for RNA Seq, microarray studies, and metabolomic analysis.  
  *Assignment*: Continue to work on class project paper.

Oct. 25  Optimizing for QPCR/Running Reactions  
  *Goal*: Run a QPCR reaction after performing necessary tests.  
  *Assignment*: Acquire QPCR data.

Nov. 1  Optimizing for QPCR/Running Reactions  
  *Goal*: Continue collecting data for class project.  
  *Assignment*: Acquire QPCR data.

Nov. 8  Analyzing QPCR data/ Finding Potential Homologues  
  *Goal*: Complete QPCR analysis for class project and related analysis. Also, introduce websites useful for finding homologous genes (Ensembl, Homologene, HUGO, Mouse Genome Informatics, etc.).  
  *Assignment*: Analyze QPCR data.

Nov. 15  Sequence Alignment  
  *Goal*: Learn to how to do pairwise and multiple sequence alignments. Discuss editing multiple sequence alignments.  
  *Assignment*: Perform pairwise and multiple sequence alignments on your project genes to prepare for phylogenetic analysis. Edit the multiple sequence alignment.

Nov. 22  Generating Phylogenies to Identify Homologues  
  *Goal*: Learn to generate neighbor-joining and maximal parsimony trees in MEGA.  
  *Assignment*: Produce a neighbor-joining tree and a maximal parsimony tree for a family of genes you are working on. Compare your genes to potential homologues in humans. Then perform a second analysis in which the genes are compared to a range of organisms to assess evolution of the gene.

* Schedule may vary subject to scheduling changes and other modifications as needed.  
Bold dates: Lab will be held in RHSC 315